

The Aging Lung

Is Lung Health Good Health for Older Adults?

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The prevalence of lung conditions, such as COPD and pulmonary fibrosis, and lung infections, such as pneumonia, increases sharply with age. The physiologic, cellular, and immunologic changes that occur during aging contribute to the development of lung disease. Studies of age-related changes in physiology and function are not only key to preventing or ameliorating disease, they are also essential for understanding healthy aging. Individuals with good lung function live longer, healthier lives, although the mechanisms by which this scenario occurs are not understood. The present article reviews changes in the aging lung that facilitate development of disease and the evidence supporting the idea that robust lung function reduces the risk of developing chronic inflammatory conditions that occur with age. CHEST 2018; ■(■):■-■

KEY WORDS: aging; COPD; idiopathic pulmonary fibrosis; immunology (lung); inflammation

Longitudinal studies on aging generally arrive at the same disturbing conclusion: the groundwork of whether a person will age well, or age poorly, begins early in life. By the time we are in our second or third decade, biological aging and chronological aging do not proceed in step.¹ Some biologic measures are more predictive of whether an individual will have a life of relatively good health, or develop chronic diseases, become frail, and die early. Intriguingly, lung function seems to play a significant role in healthy aging.^{2,3}

Physiologic changes to the lungs contribute to changes in lung function and susceptibility to disease (Table 1).⁴⁻¹¹ Lung diseases such as COPD and pulmonary fibrosis increase with age, as does the incidence of pneumonia.¹² In some cases, such as pulmonary fibrosis, the development

of disease is clearly linked to cellular senescence as mutations in genes associated with premature aging increase the risk of developing disease, and the hallmarks of cellular senescence, such as mitochondrial dysfunction, are clearly evident.¹³⁻¹⁶ In contrast to pulmonary fibrosis, mutations in genes associated with cellular senescence are not associated with an increased risk of COPD,³ and although hallmarks of cellular stress and senescence are evident once the disease is clinically apparent, these factors seem to be a consequence rather than a cause of COPD. The prevailing theory is that COPD results in age-related changes in lung capacity that are more evident in those whose initial lung capacity was low and which can be accelerated by cellular stressors such as pollution, particulates, or the unparalleled senescence-inducing properties

ABBREVIATIONS: AEC1 = alveolar epithelial type 1 cell; AEC2 = alveolar epithelial type 2 cell; ECM = extracellular matrix protein; ER = endoplasmic reticulum

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DOI: <https://doi.org/10.1016/j.chest.2018.09.003>

TABLE 1] Physiologic Changes to the Lungs and Related Changes in Lung Function and Susceptibility to Disease

Measures of Lung Function	Changes With Age	Reference
Total lung volume	No change	4, 5
Alveoli (number)	No change	6
Alveoli (size)	Increases	7
Alveoli (elasticity)	Decreases	4
Functional residual capacity The volume of air remaining at the end of expiration	Increases due to changes in elasticity and enlargement of airways	8
End-expiratory lung volume The volume of air remaining at the end of passive expiration (functional residual capacity) in addition to the increase in lung volume by applied positive end-expiratory pressure	Increases due to changes in elasticity and enlargement of airways	8
Apparent diffusion coefficient Diffusion of a gas is restricted by the boundaries of the alveoli	Increases due to airspace enlargement	7
FEV ₁ /FVC The ratio of FEV ₁ over the FVC (the amount of air that can be forcibly exhaled after taking the deepest breath possible)	Decreases due to changes in elasticity, airspace enlargement, and other physiologic changes (eg, body mass composition) that occur with age	9, 10
Airsphere wall surface area per unit volume of lung tissue	Decreases due to airspace enlargement	11

of cigarette smoke.^{17,18} Although the role of cellular senescence in COPD is not as clear, prematurely senescent mice develop features (eg, pronounced airspace enlargement) that normally would only be observed in extreme old age. This finding provides further evidence that cellular senescence contributes to both age- and disease-related changes in the lungs.^{19,20} Immunosenescence is also an important feature of the aging lung. A significant portion of the cells of the lung are resident immune cells, and the near-constant contact with circulating immune cells means that immunosenescence affects lung aging and disease progression. The present article reviews age-related changes in the lung and discusses how these factors might contribute to both disease and healthy aging (Fig 1).

Physiologic Changes in the Aging Lung

Although the number of alveoli, alveolar ducts, and capillary segments are stable once adulthood is reached⁶

and total lung volume remains the same,^{4,5} physiologic changes in the aging lung occur that decrease functional capacity. For example, alveolar and alveolar-capillary surface area increases⁶ while elasticity decreases,⁴ resulting in an increase in resting functional residual capacity and an increase in end-expiratory lung volume.⁸ In healthy older adults, these functional changes may only be felt during exercise or extreme exertion,^{21,22} but age-related changes in capacity must be accounted for when diagnosing lung disease.

Determining what “normal” lung function is in older adults has proven challenging. Some measurements of lung function such as airspace wall surface area per unit volume of lung tissue (millimeters squared per cubic millimeters) appear to decrease in a linear manner with age and be fairly consistent between the sexes.¹¹ Other measures such as the apparent diffusion coefficient, which increases as the alveoli become larger, seem to grow in a linear manner with age. These changes are not uniform over the entire lung, however, with diffusion being highest in the larger alveoli of the apical lung and

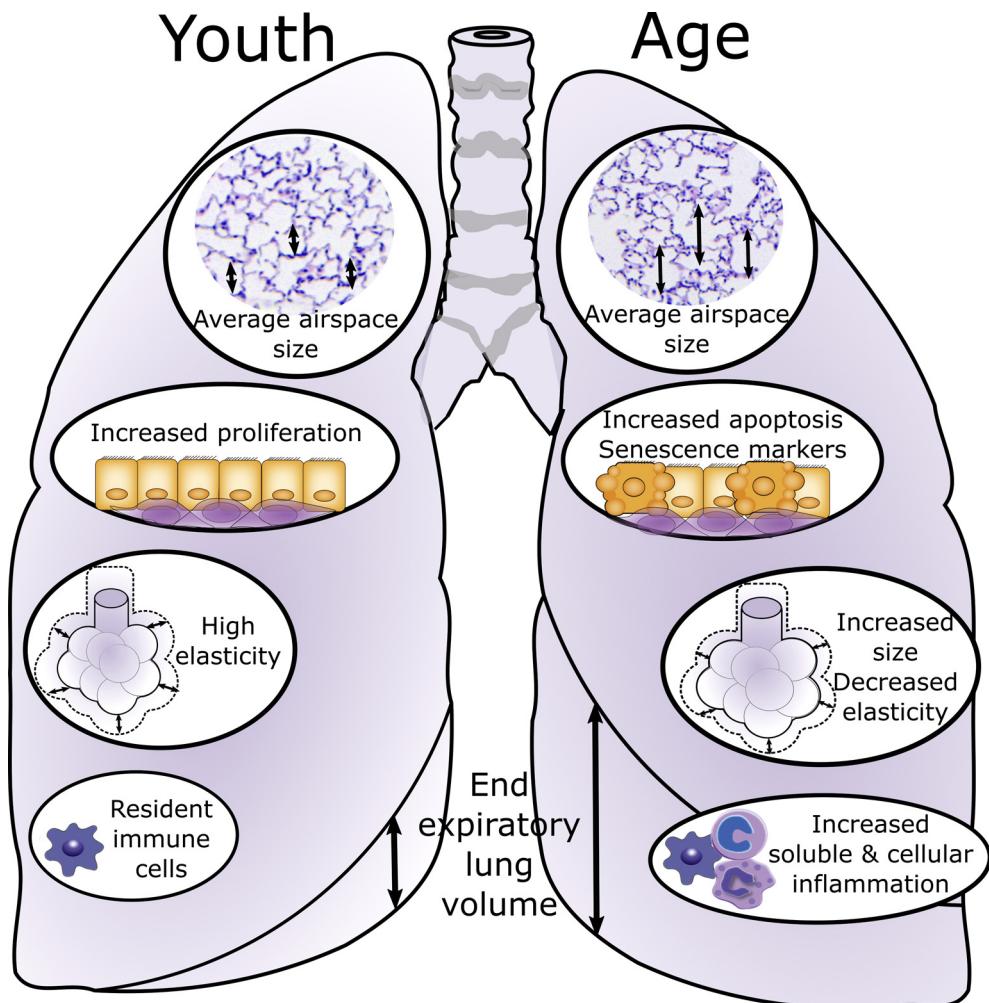


Figure 1 – Changes in the aging lung. With age, physiologic changes such as airspace enlargement and decreased elasticity of the alveoli collectively contribute to reduced forced and end-expiratory lung volume. Concurrently, increased cellular senescence of fibroblasts and epithelial cells results in reduced resilience to injury and predisposes to fibrotic scarring. Age-associated inflammation includes elevated levels of inflammatory mediators and cytokines (soluble inflammation), as well as increased numbers of immune cells such as neutrophils, even in the absence of infection (cellular inflammation). Resident immune cells such as macrophages are less effective at resolving infections and may contribute to lung remodeling.

lowest in the basal region of the lung.⁷ This scenario indicates that although there may be global increases in apparent diffusion coefficient with age, the lung does not age uniformly.

In contrast, developing reference intervals for some age-related changes such as FEV₁ and FVC have been problematic. There is some controversy as to whether FVC (the amount of air that can be forcibly exhaled after taking the deepest breath possible) and FEV₁, or the ratio of FEV₁/FVC, change in a linear manner with age; the general consensus, however, is that reference values inferred from younger populations are inaccurate when applied to older adults and can lead to a misdiagnosis of COPD.^{9,23,24} Reference values for FVC and FEV₁ have been challenging to establish due to differences attributable to height, weight, sex, and ethnicity.^{9,25,26}

This issue is further compounded when studying older adults as the relative contribution of height, weight (or more specifically, body composition), and sex to lung capacity seems to be different in older (aged > 65 years) adults compared with younger adults.^{26,27} In some studies, “normal” values seem to increase slightly higher in the “oldest old” (ie, those aged > 85 years).²⁸ As with many studies in aging, this finding is likely attributed to the “survivor effect” wherein only those with the most physiologic reserves survive past age 85 years in good enough health to be included in such studies.

Cellular Changes in the Aging Lung

The list of insults our lungs will face over the course of an average life includes particulates, ozone, aerosols, infections (and possibly overexuberant immune

responses), allergens, and pollutants. Some people will have longer lists that will include, by choice or by chance, cigarette smoke, radiation, drugs and medications, mechanical injury, and exposure to industrial pollutants. Consequently, tissue repair is of paramount importance in the lungs as we age. Inappropriate or ineffective repair can lead to scarring (ie, fibrosis/dysplasia) or remodeling (eg, emphysema, COPD). Tissue repair and remodeling are orchestrated by epithelial cells, immune cells, fibroblasts, and progenitor cells,²⁹ all of which undergo age-related changes that affect function.

Alveolar Epithelial Cells

The alveolar epithelium consists of alveolar epithelial type 1 cells (AEC1s) and type 2 cells (AEC2s), although the relative proportion of these cells is the subject of debate.³⁰⁻³³ Squamous AEC1s are flat and constitute approximately 95% of the surface area of the lung but account for a minor proportion of the total cell population. AEC1s closely interact with the alveolar capillary system and are the primary site for gas exchange. AEC2s are believed to be the long-lived progenitors of AEC1s that also produce surfactants.³⁴ Minor insults to the lung epithelium will result in destruction of AEC1s with subsequent re-epithelialization by AEC2; however, if these insults are not resolved, epithelial cells may experience endoplasmic reticulum (ER) stress which results in accumulation of immune cells that, in the best-case scenario, stimulate apoptotic cell death of the stressed epithelial cell and replenishment by AEC2s. In the worst-case scenario, aberrant or excessive immune involvement can create a cytokine environment that favors fibroblast proliferation and subsequent fibrosis or remodeling that leads to airway expansion.²⁹

Age-related changes in AECs, in conjunction with changes in the cytokine environment of the aging lung, likely contribute to susceptibility to lung infection as well as dysregulated repair responses.²⁹ Senescent epithelial cells bind more *Streptococcus pneumoniae* due to increased expression of proteins that the bacteria co-opt for cell entry³⁵; however, despite higher bacterial burdens, inflammatory signaling is muted in the lungs of aged mice.³⁶ This increased propensity to bind bacteria coupled with changes in epithelial turnover is believed to contribute to the slower rate of resolution of infection observed in older adults recovering from pneumonia.

The ratio of proliferating to apoptotic AECs seems to decrease with age, even in the absence of insult or

injury,³⁷ which seems to favor apoptotic responses and higher levels of ER stress subsequent to challenge with fibrogenic agents.³⁸ There is less evidence of ER stress in the aging lung in the absence of disease,³⁹ but there is ample evidence of ER stress in structural or immune cells in the context of COPD or pulmonary fibrosis.^{38,40,41} Once lung disease (ie, COPD, pulmonary fibrosis, lung cancer) is present, elevated levels of the senescence marker p16^{INK4a} and shorter telomere length, a cardinal sign of cellular senescence, is apparent in AECs.^{34,42} Exogenous insults such as bleomycin and cigarette smoke increase expression of these markers,⁴²⁻⁴⁵ and the degree of AEC senescence correlates with lung function⁴²; however, the degree to which AEC senescence contributes to disease is not clear. Senescence-accelerated mice developed more pronounced bleomycin-induced fibrosis,⁴⁶ implying that senescence contributes to fibrosis. In contrast, ablating senescent epithelial cells in mice exposed to cigarette smoke does not blunt the development of a COPD/emphysema type phenotype.⁴⁷ Further research disentangling the role of cellular senescence in the initiation or acceleration of lung disease is warranted.

Fibroblasts

Fibroblasts are found throughout the interstitium of the lungs and between epithelial and endothelial layers. They support the growth of epithelial cells through secretion of extracellular matrix protein (ECM). Changes in secretion of collagen contribute to age-related changes in elasticity and airspace enlargement (as reviewed by Skloot⁴⁸). During disease or experimental insult, fibroblasts from older individuals or mice produce more collagen, fibronectin, and matrix metalloproteinases, which contribute to the severity of disease^{49,50} and display all the hallmark signs of senescence (eg, telomere shortening, expression of senescence markers, decreased proliferative ability).^{15,51}

The cross-talk between cells and structure has been elegantly shown in studies comparing acellularized lungs (ie, those in which all the cells have been denuded but the ECM remains) from patients with idiopathic pulmonary fibrosis and healthy control subjects. Fibroblasts seeding fibrotic lungs take on a pro-fibrotic phenotype by expressing more α -smooth muscle actin, collagen, laminin, and fibronectin.⁵² In the fibrotic lung, structure clearly dictates function.

Although the evidence of cellular senescence in pulmonary fibrosis and other age-related lung disease is unequivocal, it is not clear to what degree alterations in

fibroblast localization, function, or ECM production change during the course of aging but in the absence of disease. Fibroblasts that have been rendered senescent in vitro have been shown to alter epithelial cell characteristics and modify lung morphology in an organoid culture system,⁵³ which presumably occurs in vivo as well. Because age is the major risk factor for lung disease, it makes intuitive sense that subtle changes in fibroblast senescence and ECM production would contribute to both age-related changes in lung elasticity and function and development of disease; however, evidence of age-related fibroblast senescence is not usually observed in the absence of experimental insult or disease.^{49,50}

Leukocytes

Resident, recruited, and transient immune cells contribute to a significant percentage of the total cells in the lung.⁵⁴ Leukocytes follow subtly different aging trajectories due to differences in origin (embryonic or bone marrow-derived), lineage (lymphoid or myeloid), and immune experience. Immunosenescence of specific immune populations in the aging lung has been reviewed in detail elsewhere.⁵⁵⁻⁵⁷ Collectively, it is apparent that immunosenescence contributes to susceptibility to infection, to increased damage during insults, and to impaired or aberrant wound healing. One underappreciated aspect of immunosenescence is an increase in systemic inflammation. Age-associated inflammation or “inflamm-aging” is the gradual increase in inflammatory cytokines in the circulation and tissues.⁵⁸ Elevated levels of cytokines and innate cells such as neutrophils are found in the lungs of older adults and aged mice,⁵⁹ but these cells are less functional due to chronic exposure to inflammatory cytokines.^{36,60-62} This immune infiltration may result from signals from senescent structural cells. When epithelial cells, specifically AEC2s, overexpress tumor necrosis factor, mice develop a pulmonary fibrosis-like phenotype with increased deposition of collagen, enlargement of airspaces, and, importantly, increased recruitment of T cells. This outcome implies that in addition to a primary insult that might induce fibrosis, a gradual increase in immune infiltration and remodeling due to age-associated inflammation contributes to lung remodeling.⁶³

Immunosenescence and age-associated inflammation contribute to both susceptibility to and severity of influenza and pneumonia. With age, the ratio of naive vs terminally differentiated T cells decreases, as does the

ability to mount robust antibody responses by B cells, which collectively contribute to susceptibility to infections.⁶⁴ Having higher than age-average levels of inflammatory cytokines in the circulation increases the risk of developing pneumonia in older adults.⁶⁵ Although a robust inflammatory response generally protects against infections, high levels of circulating inflammatory cytokines during pneumonia in the elderly are associated with more severe disease and higher mortality.⁶⁶ In fact, a meta-analysis reported that combining antibiotics with immunosuppressive steroids improved outcomes in elderly patients hospitalized for pneumonia.⁶⁷ Chronic exposure to low levels of cytokines, and especially tumor necrosis factor alpha, alters the development, migratory potential, and responsiveness of monocytes and macrophages to microbe-associated molecular patterns.^{36,60,61} Lung macrophages have alterations in their ability to regulate inflammation, which can result in hyporesponsiveness to infection⁶⁸ or hyperresponsiveness to lung injury.⁶⁹

Although the degree to which age-related changes in the lung ECM contribute to changes in leukocyte numbers or function is not clear, any detrimental age-related changes are likely compounded by those in the peripheral immune system. As an example, older mice have more fibrocytes in the bone marrow, and more of these are recruited to the lung during a bleomycin challenge,⁴⁹ thus mimicking observations observed in patients with fibrosis.⁷⁰ Studies in mice show that recruited inflammatory monocytes, which increase in number and have enhanced recruitment to sites of inflammation with age,⁶¹ contribute to the pathology of fibrosis more than resident macrophages.^{71,72} Circulating innate cells such as monocytes and macrophages seem to be particularly sensitive to the effects of age-associated inflammation,⁷³ which alters their development, phenotype, and function. As a result, increasing numbers of less mature myeloid cells enter the circulation and have hyperinflammatory responses when they encounter inflammatory stimuli, yet have altered chemotaxis and phagocytosis.^{61,62,74,75} These impairments are exaggerated in older adults with lung disease.⁷⁶⁻⁸⁰

The Relationship Between Lung Health and Other Age-Related Diseases

Maximum lung capacity is generally reached in one's mid-twenties and remains stable for a decade, or in a fortunate few, for 2 decades. Lung capacity begins to decline in mid-life. This decline is steeper for those with lung disease and for smokers; however, even among

those who have never smoked, there is considerable variation in both maximal lung capacity and the subsequent rate of decline.^{81,82} The observation that having above-average lung capacity is a predictor of good health and that lower-than-average lung capacity is a harbinger of poor health or premature death is attributed to the inventor of the spirometer, John Hutchison, in 1846⁸³ and has been confirmed many times since then.^{2,3,84-86} Intriguingly, individuals whose FEV₁ and FVC are at the lower end of the normal range develop many of the “diseases of age” (eg, cardiovascular disease, type 2 diabetes, cognitive decline) earlier than those with more robust lung function. As an example, in one longitudinal study, individuals who developed type 2 diabetes had lower FEV₁ and FVC more than a decade before they developed clinically apparent diabetes.⁸⁷ This observation that decreased lung function precedes development of diabetes (or, in some cases, even elevated blood glucose levels) has been replicated in cross-sectional^{88,89} and longitudinal^{88,90-92} studies and is independent of smoking, age, and sex. It is important to note that it is not the rate of decline that differs between those who will go on to develop diabetes and those who will not but rather their lung function when they are apparently healthy. Although a number of explanations between lung function and metabolic dysregulation have been proposed, there is no clear mechanism linking insulin resistance or hyperglycemia to lung capacity or function.^{88,93,94} A plausible explanation for the observation that lower lung capacity and function precedes type 2 diabetes is that lower lung function is an accelerant in the aging process.

Type 2 diabetes is one of the common “diseases of age,” as are cardiovascular disease, autoimmunity, and dementia. The risk of having one of these chronic conditions increases the risk, sometimes dramatically, of developing another chronic condition.⁹⁵ Only a small percentage of individuals in their forties will have more than one chronic condition but almost everyone in their eighties will have more than one.⁹⁶ The rate at which we develop these conditions is proportionate to our rate of biological aging; thus, if low lung function is an accelerant to the aging process, it should be a predictor of developing any of these age-related conditions. Consistent with this scenario, a study of middle-aged (40-70 years of age) adults found that lower FEV₁ and FVC at baseline correlated with lower cognitive function and an increased risk of being hospitalized for dementia in the 10-year follow-up period.⁹⁷ As with the studies linking lower lung function to type 2 diabetes, there was

no change in the rate of decline in lung function between the cognitively intact vs those with impairment. Other longitudinal studies have confirmed that robust lung function in mid-life correlates with higher cognitive ability later in life.^{98,99}

One of the major predictors of whether someone ages well or poorly is their level of what is called “age-associated inflammation.” With age, levels of inflammatory markers in the serum and tissues increase, and individuals with higher-than-age-average levels have higher all-cause mortality than those with lower-than-age-average levels.¹⁰⁰ The genesis of age-associated inflammation is the topic of some debate, but irrespective of the cause, it is well documented that those in the top tertile of age-associated inflammation have higher rates of chronic inflammatory conditions and are at increased risk of developing lung diseases and pneumonia.¹⁰¹ Intriguingly, lung function and age-associated inflammation seem to be inversely correlated. Common clinical measures of age-associated inflammation (eg, C-reactive protein, fibrinogen) correlate strongly with FEV₁ and FVC.¹⁰²⁻¹⁰⁴ There are conflicting reports on whether increased levels of systemic inflammation are predictive of declining lung function^{105,106} or whether they are merely higher in individuals with lower lung capacity.^{107,108} Consistent with a role of inflammation as a driver of declining lung function, there is a strong correlation between having higher-than-age-average levels of inflammatory cytokines and developing respiratory problems such as shortness of breath.¹⁰⁹

In contrast, it has been proposed that lower lung function may precede systemic inflammation. Although this theory has not been rigorously tested, longitudinal studies have found correlations between lung function and levels of inflammatory markers at a follow-up visit.^{103,104} If having low lung capacity promotes systemic inflammation, then individuals with low lung function should be at increased risk of developing other chronic inflammatory diseases. Unsurprisingly, low FVC is associated with increased cardiovascular events,¹⁰³ but surprisingly, it also precedes development of other age-related conditions such as osteoporosis¹¹⁰ and cognitive decline.¹¹¹ Consistent with the idea that inflammation drives lower lung function, even children with inflammatory diseases such as juvenile arthritis or type 1 diabetes have lower lung function.^{112,113} Although further longitudinal studies are required to determine the directionality of this relationship, it is well

documented that in individuals with COPD, levels of inflammation are higher and proportionate to disease severity,¹¹⁴ and that individuals with COPD develop other age-related conditions and cognitive decline faster than their disease-free counterparts.¹¹⁵⁻¹¹⁷

How age-associated inflammation impairs lung function (or vice versa) is not known; however, the lungs are exposed to a steady stream of microbes, pollutants, dust, and allergens. With age, innate immune cells tend to mount hyperinflammatory responses to microbes and microbial components that are slower to resolve than in younger individuals.⁶¹ Conceivably, this process could contribute to the development of age-associated inflammation. The liver usually clears microbial products that emanate from the gut; blood from the lungs, however, enters the circulation directly, and any microbial products or inflammatory mediators therein will be dispersed in the central circulation. In addition to longitudinal studies in humans, mouse models in which age-associated inflammation can be either genetically or pharmacologically enhanced or ablated⁶⁰ will be required to understand causality.

Lung Health Is Good Health for Older Adults

Lung health is intimately associated with good health in older adults. Robust lung function correlates with a higher basal metabolic rate, which is associated with keeping trim in our later years,¹¹⁸ and with activity and physical performance.¹¹⁹ As discussed earlier, having lung disease or pneumonia can accelerate the development of other, seemingly unrelated chronic conditions. Collectively, the data are clear—healthy lungs contribute to a long and healthy life—but how can we capitalize on this information? Although the data on which modifiable risk factors contribute to health and longevity are relatively simple (avoid smoke, eat well, exercise, manage any chronic conditions as well as you are able, get enough sleep, and maintain a robust social network to stave off loneliness), adherence is not as high as it should be. Perhaps by changing the messaging to include specific health advice on the role of healthy lungs in staving off some of the most feared conditions of aging while building communities that are conducive to the simplest and least expensive way to improve lung health (eg, walking) are the way to add “life to our years, rather than years to our life.”^{120,121}

Acknowledgments

Financial/nonfinancial disclosures: The author has reported to CHEST the following: D. M. E. B. is the Canada Research Chair in Aging and Immunity and is funded by the Canadian Institutes of

Health Research, the Natural Sciences and Engineering Research Council of Canada, and the The Lung Association - Ontario. Research in the Bowdish laboratory is supported by the McMaster Immunology Research Centre and the M. G. DeGroote Institute for Infectious Disease Research.

Other contributions: The author thanks Martin Stämpfli, PhD, and MyLinh Duong, FRACP, for constructive input, and Mark McDermott, PhD, for critical review of the manuscript.

References

- Belsky DW, Caspi A, Houts R, et al. Quantification of biological aging in young adults. *Proc Natl Acad Sci U S A.* 2015;112(30):E4104-E4110.
- Beatty TH, Cohen BH, Newill CA, et al. Impaired pulmonary function as a risk factor for mortality. *Am J Epidemiol.* 1982;116:102-113.
- Beatty TH, Newill CA, Cohen BH, et al. Effects of pulmonary function on mortality. *J Chron Dis.* 1985;38:703-710.
- Turner JM, Mead J, Wohl ME. Elasticity of human lungs in relation to age. *J Appl Physiol.* 1968;25:664-671.
- Briscoe WA, Dubois AB. The relationship between airway resistance, airway conductance and lung volume in subjects of different age and body size. *J Clin Invest.* 1958;37:1279-1285.
- Weibel ER, Gomez DM. Architecture of the human lung. Use of quantitative methods establishes fundamental relations between size and number of lung structures. *Science.* 1962;137:577-585.
- Fain SB, Altes TA, Panth SR, et al. Detection of age-dependent changes in healthy adult lungs with diffusion-weighted ³He MRI. *Acad Radiol.* 2005;12:1385-1393.
- DeLorey DS, Babb TG. Progressive mechanical ventilatory constraints with aging. *Am J Respir Crit Care Med.* 1999;160:169-177.
- Enright PL, Arnold A, Manolio TA, et al. Spirometry reference values for healthy elderly blacks. The Cardiovascular Health Study Research Group. *Chest.* 1996;110:1416-1424.
- Garcia-Rio F, Pino JM, Dorgham A, et al. Spirometric reference equations for European females and males aged 65-85 yrs. *Eur Respir J.* 2004;24:397-405.
- Gillooly M, Lamb D. Airspace size in lungs of lifelong non-smokers: effect of age and sex. *Thorax.* 1993;48:39-43.
- Budinger GRS, Kohanski RA, Gan W, et al. The intersection of aging biology and the pathobiology of lung diseases: a joint NHLBI/NIA workshop. *J Gerontol A Biol Sci Med Sci.* 2017;72:1492-1500.
- Coghlan MA, Shifren A, Huang HJ, et al. Sequencing of idiopathic pulmonary fibrosis-related genes reveals independent single gene associations. *BMJ Open Respir Res.* 2014;1:e000057.
- Mora AL, Rojas M, Pardo A, et al. Emerging therapies for idiopathic pulmonary fibrosis, a progressive age-related disease. *Nat Rev Drug Discov.* 2017;16:755-772.
- Alvarez D, Cardenes N, Sellares J, et al. IPF lung fibroblasts have a senescent phenotype. *Am J Physiol Lung Cell Mol Physiol.* 2017;313:L1164-L1173.
- Mora AL, Bueno M, Rojas M. Mitochondria in the spotlight of aging and idiopathic pulmonary fibrosis. *J Clin Invest.* 2017;127:405-414.
- Postma DS, Bush A, van den Berge M. Risk factors and early origins of chronic obstructive pulmonary disease. *Lancet.* 2015;385:899-909.
- Tuder RM. Aging and cigarette smoke: fueling the fire. *Am J Respir Crit Care Med.* 2006;174:490-491.
- Ishii M, Yamaguchi Y, Yamamoto H, et al. Airspace enlargement with airway cell apoptosis in Klotho mice: a model of aging lung. *J Gerontol A Biol Sci Med Sci.* 2008;63:1289-1298.
- Kurozumi M, Matsushita T, Hosokawa M, et al. Age-related changes in lung structure and function in the senescence-accelerated mouse (SAM): SAM-P1 as a new murine model of

- senile hyperinflation of lung. *Am J Respir Crit Care Med.* 1994;149:776-782.
21. Babb TG. Ventilatory response to exercise in subjects breathing CO₂ or HeO₂. *J Appl Physiol.* 1997;82:746-754.
 22. Johnson BD, Dempsey JA. Demand vs. capacity in the aging pulmonary system. *Exerc Sport Sci Rev.* 1991;19:171-210.
 23. Garcia-Rio F, Dorgham A, Pino JM, et al. Lung volume reference values for women and men 65 to 85 years of age. *Am J Respir Crit Care Med.* 2009;180:1083-1091.
 24. Hardie JA, Buist AS, Vollmer WM, et al. Risk of over-diagnosis of COPD in asymptomatic elderly never-smokers. *Eur Respir J.* 2002;20:1117-1122.
 25. Hankinson JL, Kawut SM, Shahar E, et al. Performance of American Thoracic Society-recommended spirometry reference values in a multiethnic sample of adults: the Multi-Ethnic Study of Atherosclerosis (MESA) lung study. *Chest.* 2010;137:138-145.
 26. Quanjer PH, Hall GL, Stanojevic S, et al. Age- and height-based prediction bias in spirometry reference equations. *Eur Respir J.* 2012;40:190-197.
 27. Quanjer PH, Stanojevic S, Cole TJ, et al. Multi-ethnic reference values for spirometry for the 3-95-yr age range: the global lung function 2012 equations. *Eur Respir J.* 2012;40:1324-1343.
 28. Hankinson JL, Odencrantz JR, Fedan KB. Spirometric reference values from a sample of the general US population. *Am J Respir Crit Care Med.* 1999;159:179-187.
 29. Beers MF, Morrisey EE. The three R's of lung health and disease: repair, remodeling, and regeneration. *J Clin Invest.* 2011;121:2065-2073.
 30. Lee JH, Kim J, Gludish D, et al. Surfactant protein-C chromatin-bound green fluorescence protein reporter mice reveal heterogeneity of surfactant protein C-expressing lung cells. *Am J Respir Cell Mol Biol.* 2013;48:288-298.
 31. Bantikassegn A, Song X, Politi K. Isolation of epithelial, endothelial, and immune cells from lungs of transgenic mice with oncogene-induced lung adenocarcinomas. *Am J Respir Cell Mol Biol.* 2015;52:409-417.
 32. Roper JM, Staversky RJ, Finkelstein JN, et al. Identification and isolation of mouse type II cells on the basis of intrinsic expression of enhanced green fluorescent protein. *Am J Physiol Lung Cell Mol Physiol.* 2003;285:L691-L700.
 33. Harrison JH Jr, Porretta CP, Leming K. Purification of murine pulmonary type II cells for flow cytometric cell cycle analysis. *Exp Lung Res.* 1995;21:407-421.
 34. Barkauskas CE, Cronce MJ, Rackley CR, et al. Type 2 alveolar cells are stem cells in adult lung. *J Clin Invest.* 2013;123:3025-3036.
 35. Shivshankar P, Boyd AR, Le Saux CJ, et al. Cellular senescence increases expression of bacterial ligands in the lungs and is positively correlated with increased susceptibility to pneumococcal pneumonia. *Aging Cell.* 2011;10:798-806.
 36. Hinojosa E, Boyd AR, Orihuela CJ. Age-associated inflammation and toll-like receptor dysfunction prime the lungs for pneumococcal pneumonia. *J Infect Dis.* 2009;200:546-554.
 37. Ortega-Martinez M, Rodriguez-Flores LE, Añcer-Arellano A, et al. Analysis of cell turnover in the bronchiolar epithelium through the normal aging process. *Lung.* 2016;194:581-587.
 38. Torres-Gonzalez E, Bueno M, Tanaka A, et al. Role of endoplasmic reticulum stress in age-related susceptibility to lung fibrosis. *Am J Respir Cell Mol Biol.* 2012;46:748-756.
 39. Bodas M, Min T, Vij N. Early-age-related changes in proteostasis augment immunopathogenesis of sepsis and acute lung injury. *PLoS One.* 2010;5:e15480.
 40. Min T, Bodas M, Mazur S, et al. Critical role of proteostasis imbalance in pathogenesis of COPD and severe emphysema. *J Mol Med (Berl).* 2011;89:577-593.
 41. Lawson WE, Crossno PF, Polosukhin VV, et al. Endoplasmic reticulum stress in alveolar epithelial cells is prominent in IPF: association with altered surfactant protein processing and herpesvirus infection. *Am J Physiol Lung Cell Mol Physiol.* 2008;294:L1119-L1126.
 42. Tsuji T, Aoshiba K, Nagai A. Alveolar cell senescence in patients with pulmonary emphysema. *Am J Respir Crit Care Med.* 2006;174:886-893.
 43. Aoshiba K, Tsuji T, Nagai A. Bleomycin induces cellular senescence in alveolar epithelial cells. *Eur Respir J.* 2003;22:436-443.
 44. Tsuji T, Aoshiba K, Nagai A. Cigarette smoke induces senescence in alveolar epithelial cells. *Am J Respir Cell Mol Biol.* 2004;31:643-649.
 45. Walters MS, De BP, Salit J, et al. Smoking accelerates aging of the small airway epithelium. *Respir Res.* 2014;15:94.
 46. Xu J, Gonzalez ET, Iyer SS, et al. Use of senescence-accelerated mouse model in bleomycin-induced lung injury suggests that bone marrow-derived cells can alter the outcome of lung injury in aged mice. *J Gerontol A Biol Sci Med Sci.* 2009;64:731-739.
 47. Sundar IK, Rashid K, Gerloff J, Li D, Rahman I. Genetic ablation of p16^{INK4a} does not protect against cellular senescence in mouse models of chronic obstructive pulmonary disease/emphysema. *Am J Respir Cell Mol Biol.* 2018;59(2):189-199.
 48. Skloot GS. The effects of aging on lung structure and function. *Clinics Geriatr Med.* 2017;33:447-457.
 49. Sueblinvong V, Neveu WA, Neujahr DC, et al. Aging promotes pro-fibrotic matrix production and increases fibrocyte recruitment during acute lung injury. *Adv Biosci Biotechnol.* 2014;5:19-30.
 50. Sueblinvong V, Neujahr DC, Mills ST, et al. Predisposition for disrepair in the aged lung. *Am J Med Sci.* 2012;344:41-51.
 51. Yanai H, Shtenberg A, Porat Z, et al. Cellular senescence-like features of lung fibroblasts derived from idiopathic pulmonary fibrosis patients. *Aging.* 2015;7:664-672.
 52. Booth AJ, Hadley R, Cornett AM, et al. Acellular normal and fibrotic human lung matrices as a culture system for in vitro investigation. *Am J Respir Crit Care Med.* 2012;186:866-876.
 53. Parrinello S, Coppe JP, Krtolica A, et al. Stromal-epithelial interactions in aging and cancer: senescent fibroblasts alter epithelial cell differentiation. *J Cell Sci.* 2005;118:485-496.
 54. Barletta KE, Cagnina RE, Wallace KL, et al. Leukocyte compartments in the mouse lung: distinguishing between marginated, interstitial, and alveolar cells in response to injury. *J Immunol Methods.* 2012;375:100-110.
 55. McElhaney JE, Zhou X, Talbot HK, et al. The unmet need in the elderly: how immunosenescence, CMV infection, co-morbidities and frailty are a challenge for the development of more effective influenza vaccines. *Vaccine.* 2012;30:2060-2067.
 56. Linton PJ, Thoman ML. Immunosenescence in monocytes, macrophages, and dendritic cells: lessons learned from the lung and heart. *Immunol Lett.* 2014;162:290-297.
 57. Morales-Nebreda L, Misharin AV, Perlman H, et al. The heterogeneity of lung macrophages in the susceptibility to disease. *Eur Respir Rev.* 2015;24:505-509.
 58. Claudio F, Massimiliano B, Silvana V, et al. Inflamm-aging: an evolutionary perspective on immunosenescence. *Ann N Y Acad Sci.* 2000;908:244-254.
 59. Meyer KC, Rosenthal NS, Soergel P, et al. Neutrophils and low-grade inflammation in the seemingly normal aging human lung. *Mech Ageing Dev.* 1998;104:169-181.
 60. Thevarajan N, Puchta A, Schulz C, et al. Age-associated microbial dysbiosis promotes intestinal permeability, systemic inflammation, and macrophage dysfunction. *Cell Host Microbe.* 2017;21:455-466.e454.
 61. Puchta A, Naidoo A, Verschoor CP, et al. TNF drives monocyte dysfunction with age and results in impaired anti-pneumococcal immunity. *PLoS Pathogens.* 2016;12:e1005368.
 62. Verschoor CP, Loukov D, Naidoo A, et al. Circulating TNF and mitochondrial DNA are major determinants of neutrophil phenotype in the advanced-age, frail elderly. *Mol Immunol.* 2015;65:148-156.
 63. Miyazaki Y, Araki K, Vesin C, et al. Expression of a tumor necrosis factor-alpha transgene in murine lung causes lymphocytic and fibrosing alveolitis. A mouse model of progressive pulmonary fibrosis. *J Clin Invest.* 1995;96:250-259.

64. Haq K, McElhaney JE. Immunosenescence: influenza vaccination and the elderly. *Curr Opin Immunol.* 2014;29:38-42.
65. Yende S, Alvarez K, Loehr L, et al. Epidemiology and long-term clinical and biologic risk factors for pneumonia in community-dwelling older Americans: analysis of three cohorts. *Chest.* 2013;144:1008-1017.
66. Paats MS, Bergen IM, Hanselaar WE, et al. Local and systemic cytokine profiles in nonsevere and severe community-acquired pneumonia. *Eur Respir J.* 2013;41:1378-1385.
67. Siemieniuk RA, Meade MO, Alonso-Coello P, et al. Corticosteroid therapy for patients hospitalized with community-acquired pneumonia: a systematic review and meta-analysis. *Ann Intern Med.* 2015;163:519-528.
68. Hinojosa CA, Akula Suresh Babu R, Rahman MM, et al. Elevated A20 contributes to age-dependent macrophage dysfunction in the lungs. *Exp Gerontol.* 2014;54:58-66.
69. Brandenberger C, Kling KM, Vital M, et al. The role of pulmonary and systemic immunosenescence in acute lung injury. *Aging Dis.* 2018;9:553-565.
70. Moeller A, Gilpin SE, Ask K, et al. Circulating fibrocytes are an indicator of poor prognosis in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med.* 2009;179:588-594.
71. Misharin AV, Morales-Nebreda L, Reyfman PA, et al. Monocyte-derived alveolar macrophages drive lung fibrosis and persist in the lung over the life span. *J Exp Med.* 2017;214:2387-2404.
72. Gibbons MA, MacKinnon AC, Ramachandran P, et al. Ly6Chi monocytes direct alternatively activated profibrotic macrophage regulation of lung fibrosis. *Am J Respir Crit Care Med.* 2011;184:569-581.
73. Shaw AC, Goldstein DR, Montgomery RR. Age-dependent dysregulation of innate immunity. *Nat Rev Immunol.* 2013;13:875-887.
74. Sapey E, Greenwood H, Walton G, et al. Phosphoinositide 3-kinase inhibition restores neutrophil accuracy in the elderly: toward targeted treatments for immunosenescence. *Blood.* 2014;123:239-248.
75. Butcher SK, Chahal H, Nayak L, et al. Senescence in innate immune responses: reduced neutrophil phagocytic capacity and CD16 expression in elderly humans. *J Leukoc Biol.* 2001;70:881-886.
76. Moore BB, Fry C, Zhou Y, et al. Inflammatory leukocyte phenotypes correlate with disease progression in idiopathic pulmonary fibrosis. *Front Med.* 2014;1.
77. Sapey E, Stockley JA, Greenwood H, et al. Behavioral and structural differences in migrating peripheral neutrophils from patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2011;183:1176-1186.
78. Stockley JA, Walton GM, Lord JM, et al. Aberrant neutrophil functions in stable chronic obstructive pulmonary disease: the neutrophil as an immunotherapeutic target. *Int Immunopharmacol.* 2013;17:1211-1217.
79. Bewley MA, Budd RC, Ryan E, et al. Opsonic phagocytosis in chronic obstructive pulmonary disease is enhanced by Nrf2 agonists. *Am J Respir Crit Care Med.* 2018;198(6):739-750.
80. Bewley MA, Belchamber KB, Chana KK, et al. Differential effects of p38, MAPK, PI3K or rho kinase inhibitors on bacterial phagocytosis and efferocytosis by macrophages in COPD. *PLoS One.* 2016;11:e0163139.
81. Weiss ST, Ware JH. Overview of issues in the longitudinal analysis of respiratory data. *Am J Respir Crit Care Med.* 1996;154:S208-S211.
82. Griffith KA, Sherrill DL, Siegel EM, et al. Predictors of loss of lung function in the elderly: the Cardiovascular Health Study. *Am J Respir Crit Care Med.* 2001;163:61-68.
83. Hutchinson J. On the capacity of the lungs, and on the respiratory functions, with a view of establishing a precise and easy method of detecting disease by the spirometer. *Med Chir Trans.* 1846;29:137-252.
84. Lange P, Nyboe J, Appleyard M, et al. Spirometric findings and mortality in never-smokers. *J Clin Epidemiol.* 1990;43:867-873.
85. Hole DJ, Watt GC, Davey-Smith G, et al. Impaired lung function and mortality risk in men and women: findings from the Renfrew and Paisley prospective population study. *BMJ.* 1996;313:711-715; discussion 715-716.
86. Weiss ST, Segal MR, Sparrow D, et al. Relation of FEV1 and peripheral blood leukocyte count to total mortality. The Normative Aging Study. *Am J Epidemiol.* 1995;142:493-498; discussion 499-503.
87. Litonjua AA, Lazarus R, Sparrow D, et al. Lung function in type 2 diabetes: the Normative Aging Study. *Respir Med.* 2005;99:1583-1590.
88. Lange P, Groth S, Kastrup J, et al. Diabetes mellitus, plasma glucose and lung function in a cross-sectional population study. *Eur Respir J.* 1989;2:14-19.
89. Lawlor DA, Ebrahim S, Smith GD. Associations of measures of lung function with insulin resistance and Type 2 diabetes: findings from the British Women's Heart and Health Study. *Diabetologia.* 2004;47:195-203.
90. Davis TM, Knuiman M, Kendall P, et al. Reduced pulmonary function and its associations in type 2 diabetes: the Fremantle Diabetes Study. *Diabetes Res Clin Pract.* 2000;50:153-159.
91. Engstrom G, Janzon L. Risk of developing diabetes is inversely related to lung function: a population-based cohort study. *Diabet Med.* 2002;19:167-170.
92. Engstrom G, Hedblad B, Nilsson P, et al. Lung function, insulin resistance and incidence of cardiovascular disease: a longitudinal cohort study. *J Intern Med.* 2003;253:574-581.
93. Shah SH, Sonawane P, Nahar P, et al. Pulmonary function tests in type 2 diabetes mellitus and their association with glycemic control and duration of the disease. *Lung India.* 2013;30:108-112.
94. O'Donnell CP, Tankersley CG, Polotsky VP, et al. Leptin, obesity, and respiratory function. *Respir Physiol.* 2000;119:163-170.
95. van den Akker M, Buntinx F, Metsemakers JF, et al. Multimorbidity in general practice: prevalence, incidence, and determinants of co-occurring chronic and recurrent diseases. *J Clin Epidemiol.* 1998;51:367-375.
96. Gallacher KI, McQueenie R, Nicholl B, et al. Risk factors and mortality associated with multimorbidity in people with stroke or transient ischaemic attack: a study of 8,751 UK Biobank participants. *J Comorbid.* 2018;8:1-8.
97. Pathan SS, Gottesman RF, Mosley TH, et al. Association of lung function with cognitive decline and dementia: the Atherosclerosis Risk in Communities (ARIC) Study. *Eur J Neurol.* 2011;18:888-898.
98. Richards M, Strachan D, Hardy R, et al. Lung function and cognitive ability in a longitudinal birth cohort study. *Psychosom Med.* 2005;67:602-608.
99. Barnes DE, Yaffe K, Satariano WA, et al. A longitudinal study of cardiorespiratory fitness and cognitive function in healthy older adults. *J Am Geriatrics Soc.* 2003;51:459-465.
100. Harris TB, Ferrucci L, Tracy RP, et al. Associations of elevated interleukin-6 and C-reactive protein levels with mortality in the elderly. *Am J Med.* 1999;106:506-512.
101. Yende S, Tuomanen EI, Wunderink R, et al. Preinfection systemic inflammatory markers and risk of hospitalization due to pneumonia. *Am J Respir Crit Care Med.* 2005;172:1440-1446.
102. Yohannes AM, Tampubolon G. Changes in lung function in older people from the English Longitudinal Study of Ageing. *Expert Rev Respir Med.* 2014;8:515-521.
103. Engstrom G, Lind P, Hedblad B, et al. Lung function and cardiovascular risk: relationship with inflammation-sensitive plasma proteins. *Circulation.* 2002;106:2555-2560.
104. Bolton CE, Schumacher W, Cockcroft JR, et al. The CRP genotype, serum levels and lung function in men: the Caerphilly Prospective Study. *Clin Sci.* 2011;120:347-355.
105. Rasmussen F, Mikkelsen D, Hancox RJ, et al. High-sensitive C-reactive protein is associated with reduced lung function in young adults. *Eur Respir J.* 2009;33:382-388.

106. Shaaban R, Kony S, Driss F, et al. Change in C-reactive protein levels and FEV1 decline: a longitudinal population-based study. *Respir Med.* 2006;100:2112-2120.
107. Hancox RJ, Gray AR, Sears MR, et al. Systemic inflammation and lung function: a longitudinal analysis. *Respir Med.* 2016;111:54-59.
108. Fogarty AW, Jones S, Britton JR, et al. Systemic inflammation and decline in lung function in a general population: a prospective study. *Thorax.* 2007;62:515-520.
109. Gimeno D, Delclos GL, Ferrie JE, et al. Association of CRP and IL-6 with lung function in a middle-aged population initially free from self-reported respiratory problems: the Whitehall II study. *Eur J Epidemiol.* 2011;26:135-144.
110. Choi JW, Pai SH. Association between respiratory function and osteoporosis in pre- and postmenopausal women. *Maturitas.* 2004;48:253-258.
111. Singh-Manoux A, Dugravot A, Kauffmann F, et al. Association of lung function with physical, mental and cognitive function in early old age. *Age.* 2011;33:385-392.
112. Wagener JS, Taussig LM, DeBenedetti C, et al. Pulmonary function in juvenile rheumatoid arthritis. *J Pediatr.* 1981;99:108-110.
113. Cazzato S, Bernardi F, Salardi S, et al. Lung function in children with diabetes mellitus. *Pediatric Pulmonol.* 2004;37:17-23.
114. Broekhuizen R, Wouters EF, Creutzberg EC, et al. Raised CRP levels mark metabolic and functional impairment in advanced COPD. *Thorax.* 2006;61:17-22.
115. Incalzi RA, Caradonna P, Ranieri P, et al. Correlates of osteoporosis in chronic obstructive pulmonary disease. *Respir Med.* 2000;94:1079-1084.
116. Hung WW, Wisnivesky JP, Siu AL, et al. Cognitive decline among patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2009;180(2):134-137.
117. Ozge C, Ozge A, Unal O. Cognitive and functional deterioration in patients with severe COPD. *Behav Neurol.* 2006;17:121-130.
118. Choi JW, Pai SH. Brief communication: respiratory function is closely associated with basal metabolic rate in elderly persons. *Ann Clin Lab Sci.* 2004;34:99-102.
119. Simpson CF, Punjabi NM, Wolfenden L, et al. Relationship between lung function and physical performance in disabled older women. *J Gerontol A Biol Sci Med Sci.* 2005;60:350-354.
120. Your lungs and exercise. *Breathe (Sheff).* 2016;12(1):97-100.
121. Li F, Fisher KJ, Brownson RC, et al. Multilevel modelling of built environment characteristics related to neighbourhood walking activity in older adults. *J Epidemiol Commun Health.* 2005;59:558-564.